Study of Potential Probiotic Properties of Lactic Acid Bacteria Isolated from Raw and Traditional Fermented Camel Milk

M. Mahmoudi 1, M. Khomeiri 1*, M. Saeidi 2, M. Kashaninejad 1, and H. Davoodi 3

ABSTRACT

In the present research work, the potential probiotic properties of Lactococcus lactis KMCM3 and Lactobacillus helveticus KMCH1 isolated from raw camel milk and traditional fermented camel milk (Chal), respectively, were studied. The probiotic properties of isolates that were investigated included the hemolysis, antibiotic resistance, antibacterial features, resistance to low pH and bile salts, survival under simulated GastroIntestinal Tract (GIT) conditions, adhesion ability to hydrocarbon, and their auto-aggregation and co-aggregation rates. None of isolates exhibited hemolytic activity. They were susceptible against tetracycline, penicillin, ampicillin, chloramphenicol, erythromycin and vancomycin. Lac. lactis KMCM3 and L. helveticus KMCH1 retained their viability at pH 3.0 (8.68 and 8.6 log cfu mL⁻¹, respectively), 0.3% w/v bile salts (8.23 and 8.58 log cfu mL⁻¹, respectively) and under simulated GIT conditions (8.31 and 8.46 log cfu mL⁻¹, respectively). Both of these isolates inhibited the growth of E. coli, S. aureus, L. monocytogenes, B. cereus and S. enterica subsp. enterica serovar Typhimurium with MIC values of 6.25 to 25 mg mL⁻¹. In addition, They exhibited an ability to adhere to hydrocarbon (xylene), and possessed a high auto-aggregation and co-aggregation rate (more than 40%).

Keywords: Lactococcus lactis, Lactobacillus helveticus, Chal, Auto-aggregation rate, Co-aggregation rate

INTRODUCTION

Camel Milk (CM) and traditional Fermented Camel Milk (FCM) are widely consumed as important sources of human nutrition in Africa and the Middle Eastern countries (Fguiri et al., 2015). CM contains low amounts of cholesterol. CM can be considered as a nutritious product with high stability due to high contents of antimicrobial agents such as lysozyme, lactoperoxidase, lactoferrin, and immunoglobulin (Khalesi et al., 2017). It is easily digestible due to soft coagulum formation after milk ingestion in the gastrointestinal tract (Shamsia, 2009).

Chal is an Iranian traditional FCM; it involves spontaneous fermentation without the addition of a starter culture and is produced in Turkmen Sahra and AqQala, Golestan Province, Iran (Soleymanzadeh et al., 2016). Traditional products of FCM are consumed under different names in other countries such as Gariss in Sudan (Ashmaig et al., 2009), Susaac in Kenya (Fguiri et al., 2015), and Shubat in Kazakhstan (Akhmetsadykova et al., 2015), and Ititu in Ethiopia (Seifu et al., 2012). It is known as a functional food because of claimed health benefits such as its traditionally anti-infective, anti-cancer, antidiabetic effects (Fguiri et al., 2015). Also, Ayyash et al. (2018) reported that the proteolytic, antioxidant, anti-cancer activity and ACE-
inhibition of water soluble extracts from all FCMs were higher than those of fermented bovine milk. There has been a great interest towards the isolation of new probiotic strains with health promoting benefits in order to use these bacteria in industrial and pharmaceutical applications (Khan, 2014).

Different species of Lactic Acid Bacteria (LAB) such as \textit{L. plantarum}, \textit{L. kefiri}, \textit{L. paracasei}, \textit{L. casei}, and \textit{Enterococcus faecium} are involved in the fermentation of CM (Soleymanzadeh \textit{et al.}, 2016; Akhmetzadykova \textit{et al.}, 2015). CM and FCM are important sources for the isolation of LAB having a high probiotic potential (Abushelaibi \textit{et al.}, 2017).

However, studying or identifying the LAB isolates in traditional fermented dairy products can be useful for their application in the industrial production of functional dairy products with indigenous strains (Ashmaig \textit{et al.}, 2009). The aim of this study was to identify the LAB isolated from CM and Iranian traditional FCM (Chal), and to investigate their probiotic potential, including the antibacterial features, resistance to low pH and bile salts, survival under simulated GastroIntestinal Tract (GIT) conditions, adhesion ability to hydrocarbon, and finally, their auto-aggregation and co-aggregation rates. Thus, using these parameters, the effectiveness of the probiotic cultures can be understood for development of functional products in the dairy industry.

**MATERIALS AND METHODS**

**Isolation and Initial Screening of LAB**

Ten samples of CM and Chal were collected in sterilized bottles from Turkman Sahra and AqQala, Iran. Briefly, 10 mL of raw milk and Chal samples were added to 90 mL of sterile NaCl solution (0.85% w/v). The suspensions were homogenized with a vortex, then, 100 µL of an appropriate dilution (10^{1} to 10^{3}) was spread on MRS agar. The plates were incubated anaerobically using a gas pack system (Anaerocult A, Merck, Germany) at 37°C for 48 hours. After the incubation time, the isolated bacteria were selected based on microscopic characteristics, Gram staining, and catalase activity for molecular identification (Ashmaig \textit{et al.}, 2009).

**Molecular Identification of Isolates**

The genomic DNA extraction was performed according to the instructions of the DNA extraction kit (Yekta Tajhiz Azma, Iran). The amplification of \textit{16S rDNA} gene (1500 bp) was performed using a universal primer pair: 27F 5´-\textit{AGAGTTTGATCCTGGCTCAG} -3´ and 1492R 5´-\textit{GGTTACCTTGTTACGACTT} -3´ (Leite \textit{et al.}, 2015). The thermal cycler program used for the PCR reaction consisted of an initial denaturation at 95°C for 5 minutes; followed by 30 cycles of denaturation at 94°C for 30 seconds, annealing at 54°C for 30 seconds, extension at 72°C for 2 minutes; and then final extension at 72°C for 10 minutes. The sequencing of PCR products was carried out by Macrogen Inc. (Seoul, Korea). Finally, to identify each isolate, the sequences were analyzed using the BLAST program of the NCBI and selected based on the highest percentage of identity.

**Hemolysis**

To determine the non-pathogenic bacteria, their hemolytic activity was evaluated according to the method by Tejero-Sariñena \textit{et al.} (2012). Each isolate was cultured on a blood agar plate containing 5% sheep blood and incubated for 48 hours at 37°C. Hemolytic activity is characterized by the observation of either clear zones around the colonies (β-hemolysis), green-hued zones around the colonies (α-hemolysis), or absence of any zone around the colonies (γ-hemolysis).

**Antibiotic Resistance**

The disk diffusion method was applied to determine the antibiotic susceptibility of
isolates against commonly prescribed antibiotics such as tetracycline, penicillin, ampicillin, chloramphenicol, erythromycin, vancomycin, kanamycin, streptomycin and gentamycin (PadtanTeb Co., Iran) as reported by Vijayakumar et al. (2015). The results were interpreted as susceptible, intermediate, or resistant based on CLSI (2013).

**Tolerance to Acidic and Bile Salts Conditions**

The acid and bile salts tolerance was tested as reported by Nami et al. (2014b). The 24-hour bacterial cultures were inoculated into MRS broth adjusted to pH 3.0 or MRS broth containing 0.3% (w/v) bile salts. Suspensions were then incubated at 37°C for 2 hours under acidic conditions and for 2 and 3 h under bile salts conditions.

**Survival Assessment under Simulated GastroIntestinal Tract (GIT) Conditions**

The survival of isolates during simulated GIT passage was investigated according to the method by Nami et al. (2014b) with some modifications. Briefly, the cultures in a stationary phase were centrifuged at 4,000 rpm for 10 minutes. Then, a cell pellet of each bacterium was resuspended in 1 mL of MRS broth, then inoculated into 9 mL of MRS broth adjusted to pH 3.0 (by adding 4 N HCl) and later supplemented with a filter-sterilized solution of pepsin (3 mg mL⁻¹). The simulated gastric juice was incubated for 2 hours at 37°C. Subsequently, to create a simulated intestinal condition, a sterile solution of 4 N NaOH was added to bring the pH to 6.5, supplemented by a filter sterilized bile salt solution (0.3% w/v) and pancreatin solution (0.1% w/v). The simulated intestinal environment was incubated at 37°C and the sampling was performed at an interval of 2 and 3 hours, as these periods represent the simulation of fast and slow digestion in the intestine. The viable cell counts were done on MRS agar to determine the survival rate.

**Preparation of Cell Free Supernatant (CFS)**

Each bacterium was inoculated into a MRS broth and incubated at 37°C under anaerobic conditions to reach the end of the logarithmic phase. After centrifugation at 14,000×g for 15 minutes at 4°C, the pH of CFS was neutralized to 7.2 by adding 5N NaOH. The neutralized and acidic CFSs were sterilized by using a 0.22 μm sterile syringe filter and were frozen at -20°C followed by freeze drying. On the test day, the freeze dried CFS was reconstituted with 1 mL of deionized water (Nami et al., 2014a).

**Antibacterial Activity**

The microdilution method was used to evaluate the antimicrobial properties of LAB CFSs against pathogenic bacteria. Briefly, 180 μL of diluted CFS in Muller-Hinton Broth and 20 μL of each bacterial suspension (10⁵ cfu mL⁻¹) were added to each well. After 20 h of incubation, the lowest concentration of CFS that completely inhibited the growth of pathogenic bacteria was reported as Minimum Inhibitory Concentrations MIC. To determine the Minimum Bactericidal Concentration (MBC), 10 μL of each well of MIC was spotted on MHA and incubated at 37°C for 24 hours (Ben Slama et al., 2013).

**Auto-aggregation and Co-aggregation**

The auto-aggregation and co-aggregation ability of the isolates were evaluated according to the method described by Collado et al. (2008) with slight modifications. Briefly, bacterial suspension were prepared in PBS and adjusted to an absorbance (A) of about 0.25±0.05 at 600 nm. Then, the samples were
incubated at room temperature without agitation. The auto-aggregation percentage is calculated as $1-(A_t/A_0)\times100$, where $A_t$ represents absorbance at 600 nm at time $t=5$ h, and $A_0$ the absorbance at $t=0$ h.

For the co-aggregation assay, equal volumes of each LAB suspension and pathogenic bacteria were mixed. The percentage of co-aggregation was expressed as $1-(A_{\text{mix}})/(A_{\text{pat}}+A_{\text{pro}}/2)\times100$, where $A_{\text{pat}}$ and $A_{\text{pro}}$ represent the absorbance of each bacterial suspension in control tubes including pathogenic and probiotic bacteria at 0 hour, and $A_{\text{mix}}$ represents the absorbance of the two mixed bacterial suspensions at 5 hours.

**Cell Surface Hydrophobicity**

Briefly, equal volumes of bacterial suspension and solvent were transferred to the tube and the two-phase mixture was completely mixed. After 1 hour, the hydrophobicity was reported as: $(A_0-A)/A_0\times100$, where $A_0$ and $A$ are the Absorbance before and after separation of the aqueous phase, respectively (Collado et al., 2008).

**Statistical Analysis**

All experiments were performed in triplicates. Statistical analysis was done on the data in a randomized complete design by using the SAS 9.1.3. Software, and Duncan test was applied to compare the averages at 5% level. The values were presented as mean±standard deviation.

**RESULTS AND DISCUSSION**

**Molecular Identification of LAB Isolated From Camel Milk and Chal**

Based on the BLAST results of sequences, two bacterial isolates, namely, *Lac. lactis* KMCM3 and *L. helveticus* KMCH1 isolated from CM and Chal, respectively, with the highest percentage of identity (97%) were selected for further studies. Based on previous researches, the *Lac. lactis*, *Lac. garvieae*, *L. reuteri* and *L. plantarum* were isolated from raw CM in Abu Dhabi (United Arab Emirates) (Abushelaibiet al., 2017). *Enterococcus*, *Lactobacillus* and *Lactococcus* genera were dominant in raw and fermented CM from Kazakhstan (Akhmetsadykova et al., 2015). Leite et al. (2015) also identified 34 isolates from four Brazilian kefir grains by 16S rDNA gene sequencing. Eighteen isolates belonged to *Leuconostoc mesenteroides*, 11 to *Lactococcus lactis* and 5 to *Lactobacillus paracasei*. Also, Soleymanzadeh et al. (2016) isolated *L. kefiri*, *L. gasseri*, *L. paracasei*, *L. plantarum*, *L. paraplantarum*, *Leuconostoc lactis*, *Enterococcus faecium*, *Weissella cibaria* from Chal by 16S rRNA gene sequences. LAB species isolated from spontaneously FCM were *Lac. lactis*, *Pediococcus acidilactici*, *Lactobacillus plantarum*, *Streptococcus lutetiensis* and *Weissella confuse* which were identified through 16S rRNA gene sequencing (Fugl et al., 2017).

However, to our knowledge, no other researcher has reported the isolation of *L. helveticus* from traditional FCM. The sequencing results and the isolation origin of the bacteria are presented in Table 1.

**Hemolysis Activity**

According to the recommendation of FAO/WHO (2002), non-hemolytic activity is the first property for selection of a probiotic strain, since it indicates that the bacteria are not pathogenic. According to our results, none of the selected isolates was able to hydrolyze red blood cells when grown in blood agar. These results are in agreement with studies of Abushelaibi et al. (2017) and Tejero-Sariñena et al. (2012).

**Antibiotic Susceptibility of LAB**

According to the results presented in Table 2, *L. helveticus* KMCH1 is resistant to...
Table 1. Sequencing results of PCR products for identification of LAB species isolated from raw and traditional FCM (Chal).

<table>
<thead>
<tr>
<th>Initial identification code</th>
<th>Sequencing results</th>
<th>Identity</th>
<th>Source</th>
<th>Sample collection location</th>
</tr>
</thead>
<tbody>
<tr>
<td>KMCM3</td>
<td>Lactococcus lactis</td>
<td>97%</td>
<td>CM</td>
<td>AqQala, Iran</td>
</tr>
<tr>
<td>KMCH1</td>
<td>Lactobacillus helveticus</td>
<td>97%</td>
<td>(Chal)</td>
<td>TurkmanSahra, Iran</td>
</tr>
</tbody>
</table>

Table 2. Susceptibility of isolates to different antibiotics.

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>Concentration (μg disk⁻¹)</th>
<th>Diameter of inhibition zone (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell wall inhibitors</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Vancomycin</td>
<td>30 μg</td>
<td>18 (S)</td>
</tr>
<tr>
<td>Penicillin</td>
<td>10 μg</td>
<td>24 (S)</td>
</tr>
<tr>
<td>Ampicillin</td>
<td>10 μg</td>
<td>20 (S)</td>
</tr>
<tr>
<td>Protein synthesis inhibitors</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tetracycline</td>
<td>30 μg</td>
<td>15 (I)</td>
</tr>
<tr>
<td>Chloramphenicol</td>
<td>30 μg</td>
<td>23 (S)</td>
</tr>
<tr>
<td>Streptomycin</td>
<td>10 μg</td>
<td>0 (R)</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>10 μg</td>
<td>10 (R)</td>
</tr>
<tr>
<td>Kanamycin</td>
<td>30 μg</td>
<td>11 (R)</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>15 μg</td>
<td>19 (I)</td>
</tr>
</tbody>
</table>

a S: Susceptible, I: Intermediate, R: Resistant.

vancomycin (a glycopeptide), kanamycin, gentamycin, and streptomycin (aminoglycosides), while Lactococcus lactis KMCM1 is resistant to aminoglycosides and susceptible to the other antibiotics. Generally, the LAB isolates tested were sensitive to tetracycline, ampicillin, penicillin, chloramphenicol, and erythromycin, which are commonly administered for the treatment of gastrointestinal infections (Tejero-Sariñena et al., 2012). In particular, resistance to vancomycin is a major concern because it is one of the few antibiotics that have a broad effect against clinical infections caused by multi-drug resistant pathogens (Zhou et al., 2005). In our study, it has been observed that Lactococcus lactis KMCM3 had good sensitivity to most of the tested antibiotics, particularly vancomycin. Our results are similar to those obtained by Tejero-Sariñena et al. (2012) and Nami et al. (2014b). The resistance to vancomycin is an intrinsic property in many Lactobacillus species that could be attributed to the presence of D-Ala-D-lactate instead of the normal dipeptide D-Ala-D-Ala in their peptidoglycan (Ammor et al., 2008). The resistance to aminoglycosides is attributed to the absence of a cytochrome-mediated electron transport system that mediates in the antibiotic uptake (Argyri et al., 2013). In the intrinsic resistance to antibiotics, the resistance genes are chromosomally encoded and cannot be transmitted to other bacteria. Thus, the risk of transmission to other organisms can be minimized (Tejero-Sariñena et al., 2012).

Acid and Bile Salts Resistance

A pH value below 3.0 is not the most common pH value in the human stomach (Argyri et al., 2013). Resistance to acid stress is one of the essential properties for probiotic microorganisms. According to our
results in Figure 1-a, no significant reduction was observed in viable counts of the tested LAB at pH 3.0 at the 2 hours time-point. They maintained their survival at more than 8.0 log cfu mL\(^{-1}\). Similar results are reported by Nami et al. (2014b) and Abushelaibi et al. (2017). The acid tolerance of LAB is attributed to F0F1-ATPase activity, which is activated when the extracellular pH is low, to increase the intracellular pH (Corcoran et al., 2005).

The bile salts tolerance is an important property for survival of probiotics in the small intestine to survive and exert their health benefits in GIT (Argyri et al., 2013). Based on our findings in Figure 1-b, Lac. lactis KMCM3 showed a significant decrease (P< 0.05) in bile salts in comparison to L. helveticus KMCH1, however both of them retained their viability more than 8.0 log cfu mL\(^{-1}\). Our results are consistent with the results of Nami et al. (2014b), Lee et al. (2015), and Bian et al. (2016).

**Survival in GIT Conditions**

Screening of potential probiotic strains is based on their tolerance under GIT conditions (Vera-Pingitore et al., 2016). As shown in Figure 2, the LAB testing shows no significant difference in simulated gastric juice environment. Mahmoudi et al. (2016)
reported that the action of pepsin on the cell membrane is not lethal to most LAB. This result is in accordance with Bian et al. (2016), whereas Lee et al. (2015) reported a significant decrease, which is approximately 3.0 log cfu mL\(^{-1}\) for Lac. lactis K24 after 2 hours of incubation in simulated gastric juice. This difference may be dependent on the strain. For probiotics to exert health benefits, they should be resistant to stomach acid stress and must survive the passage through the small intestine into the large intestine for subsequent colonization (Nami et al., 2014b; Mahmoudi et al., 2016). The tested LAB indicated a significant decrease (P< 0.05) while passing through the GIT, however, they retained their survival in the range of 8.0–9.0 log cfu mL\(^{-1}\). According to our findings, the resistance of Lac. lactis KMCM3 isolated from CM was remarkably more than Lac. lactis 2HL isolated from vaginal microflora by Nami et al. (2014b). This difference could be related to the origin of isolation and strain. Based on the data obtained from bile salts and GIT tolerance tests, the significant decrease observed for L. helveticus KMCH1 may be owing to the effect of pancreatin on the cell wall or membrane components (Ferrando et al., 2016). In conclusion, the survival rate of the tested LAB was excellent in the simulated GIT and could exert the expected health benefits.

### Anti-Pathogen Features

Our results as seen in Table 3 indicate that the isolated LAB have MICs of 6.25 to 25 mg mL\(^{-1}\) against Gram-positive and Gram-negative pathogenic bacteria. This result reveals that the related MIC values of CFS of the analyzed LAB have wide spectrum of anti-pathogenic activity. Considering that the neutralized CFSs exerted no antibacterial effect (data not shown), it can be concluded that antimicrobial activity of the CFSs is related to pH. Antimicrobial activity of LAB strains may be due to the production of organic acids, bacteriocins or other metabolites (Abushelaibi et al., 2017). The effects of antimicrobial activity of L. helveticus, isolated from poultry waste (Ayantola and Oladunmoye, 2016), and L. helveticus, isolated from Sinkiang traditional cheese (Bian et al., 2016), were attributed to the production of organic acids such as lactic and acetic acid. There is a hypothesis that organic acids, by neutralizing the cytoplasmic membrane’s electrochemical potential, increase the membrane...
Table 3. Antimicrobial activity of LAB species CFSs \textsuperscript{a} against Gram-positive and Gram-negative pathogenic bacteria.

<table>
<thead>
<tr>
<th>Pathogenic bacteria</th>
<th>ATCC No.</th>
<th>MIC\textsuperscript{b} (mg mL\textsuperscript{-1})</th>
<th>MBC\textsuperscript{c} (mg mL\textsuperscript{-1})</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lac. lactis KMCM3</td>
<td>L. helveticus KMCH1</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>25922</td>
<td>12.5</td>
<td>6.25</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>25923</td>
<td>25</td>
<td>6.25</td>
</tr>
<tr>
<td>Listeria monocytogenes</td>
<td>19115</td>
<td>12.5</td>
<td>6.25</td>
</tr>
<tr>
<td>Bacillus cereus</td>
<td>11778</td>
<td>12.5</td>
<td>6.25</td>
</tr>
<tr>
<td>Salmonella enterica subsp. enterica ser. Typhimurium</td>
<td>14028</td>
<td>12.5</td>
<td>6.25</td>
</tr>
</tbody>
</table>

\textsuperscript{a} Cell Free Supernatant; \textsuperscript{b} Minimum Inhibitory Concentration, \textsuperscript{c} Minimum Bactericidal Concentration.

permeability causing bacteriostasis and, subsequently, death of the susceptible cells (Da\'li\'e \textit{et al}., 2010). Other researchers have confirmed the inhibitory activity of organic acids and bacteriocins as produced by \textit{Lac. lactis} strains (Hwanhlem \textit{et al}., 2017; Kruger \textit{et al}., 2013). The bacteriocins’ mechanism of antimicrobial action could be related to the steps-adsorption of the bacteriocin on the cell wall, its transmission through the membrane and, finally, its activity within the cytoplasm (Garcha and Sharma, 2013).

Auto-aggregation, Co-aggregation, and Hydrophobicity Abilities

Cell adhesion is the process in which cells interact and attach to interacting surfaces or another cell (Kos \textit{et al}., 2003). It has been suggested that the bacterial strains’ ability to adhere to epithelial cells and mucosal surfaces is another important property for selection of potential probiotic strains (Vijayakumar \textit{et al}., 2015). Based on several studies, it is observed that the aggregation ability is attributed to cell adherence properties (Kos \textit{et al}., 2003; Del Re \textit{et al}., 2000). The ability of microorganisms belonging to the same bacterial strain to aggregate is known as auto-aggregation; co-aggregation is associated with the aggregation between two different bacterial strains. Cell surface hydrophobicity is defined as the ability of a strain to adhere to hydrocarbons (Collado \textit{et al}., 2008). A correlation has been reported between auto-aggregation and adhesion ability in \textit{L. acidophilus} M92 (Kos \textit{et al}., 2003) as well as a relationship between adhesion and hydrophobicity (Del Re \textit{et al}., 2000) factors in some \textit{Bifidobacterium} strains.

As seen in our results in Figure 3, the tested LAB show good auto-aggregation percentages, (more than 40%), indicating that such strains have a probiotic capacity (Peres \textit{et al}., 2014). These results are different from other \textit{Lac. lactis} strains that indicated strain-dependence as reported by Abushelaibi \textit{et al}., (2017).

Both LAB isolates showed co-aggregation ability with all the tested pathogenic bacteria (Figure 4). \textit{L. helveticus} KMCH1 demonstrated a remarkable co-aggregation with all pathogens (more than 40%), but \textit{L. monocytogenes} exhibited a high co-aggregation ability with both of the tested LAB (to a level of more than 50%). Moreover, \textit{Lac. lactis} KMCM3 showed the lowest levels of co-aggregation toward \textit{salmonella enterica} subsp. enterica serovar \textit{Typhimurium}, and \textit{S. aureus} (less than 40%) (P< 0.05). The co-aggregation of food-associated LAB with pathogens is of special interest for potential applications since it involves protecting the human gut from the
Potential Probiotic of LAB from Camel Milk

Figure 3. Auto-aggregation and cell surface hydrophobicity ability of *L. helveticus* KMCH1 and *Lac. lactis* KCMCM3 as a percentage. The results are represented as Mean of triplicates±SD. Values presented are not statistically significant in the Duncan test (P> 0.05).

Figure 4. Percentage of co-aggregation of *L. helveticus* KMCH1 and *Lac. lactis* KCMCM3 with pathogens (*Escherichia coli*, *Staphylococcus aureus*, *Salmonella enterica* subsp. *Enterica* serovar *Typhimurium*, *Bacillus cereus*, and *Listeria monocytogenes*) after 5 hours co-incubation at room temperature. Bars with no common letter are statistically significant (P< 0.05) from each other; lower case letters show differences between pathogens for each *L. helveticus* KMCH1 and *Lac. lactis* KCMCM3 strain and capital letters show differences of each pathogen between *L. helveticus* KMCH1 and *Lac. lactis* KCMCM3 strains.

Colonization of pathogens and infection of the gastrointestinal tract (Peres *et al.*, 2014).

The ability of bacteria to adhere to xylene reflects the cell surface hydrophobicity or hydrophilicity (Kos *et al.*, 2003). According to Figure 3, there is no significant difference between the two tested LAB in terms of affinity to the hydrocarbon. Both showed good adhesion capabilities towards xylene, indicating the hydrophobic property of the cell surface. It has been proposed that the probiotics possessing hydrophobic cell surface property are capable of adhering to the intestinal mucosa. Although hydrophobicity may represent adhesion capability, it cannot be a prerequisite for strong adhesion to the human intestinal cells (Todorov *et al.*, 2008). From previous studies carried out on the microbial cell surface chemistry, it appears that hydrophobicity is related to the presence of (glyco-) proteinaceous material, whereas hydrophilicity is due to the presence of polysaccharides (Kos *et al.*, 2003).
CONCLUSIONS

In the present study, *Lactococcus lactis* KMCM3 and *Lactobacillus helveticus* KMCH1 isolated from camel raw milk and Chal, respectively, displayed the absence of hemolytic activity as well as sensitivity to antibiotics. Therefore, they are considered safe. Both of the LAB isolates showed a wide antibacterial activity spectrum. Furthermore, they displayed potential probiotic properties such as a remarkable survival rate under simulated GIT conditions. The tested LAB had high percentages of auto-aggregation, co-aggregation, and adhesion capabilities to the hydrocarbon xylene. These characteristics are related to the capability of isolates to adhere to the intestinal epithelial cells to compete with pathogens. Thus, we conclude that *Lactococcus lactis* KMCM3 and *Lactobacillus helveticus* KMCH1 have all the necessary probiotic properties required for use as a probiotic culture in the development of functional dairy products.

REFERENCES


15. Ferrando, V., Quiberoni, A., Reinheimer, J. and Suárez, V. 2016. Functional Properties of *Lactobacillus plantarum* Strains: A Study In


بررسی ویژگی‌های پریوتابیک بالقوه باکتری‌های لاکتیک اسیذ جذا شذه از شی خام و شیر شتر تخلمیری سنتی
م. محمودی، م. خمیری، م. سعیذی، م. کاشانی نژاد و ه. داوودی

چکیده

Lactococcus lactis در پژوهش حاضر، ویژگی‌های پریوتابیک بالقوه Lactobacillus helveticus KMCH1 و Lactobacillus plantarum KCC-24 Isolated from Italian Rye-Grass (Loliummultiflorum) Forage. Anaerobe. 32: 90-97.


Downloaded from journals.modares.ac.ir at 18:08 IRST on Saturday October 26th 2019